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Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

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 $\label{thm:continuous} \textbf{To cite this Article } \ Christodoulou, \ Chris \ , \ Agrawal, \ Sudhir \ and \ Gait, \ Michael \ J. (1987) \ 'A \ New 5'-Protecting \ Group \ for \ Use in Solid-Phase \ Synthesis \ of \ Oligoribonucleotides', \ Nucleotides \ and \ Nucleic \ Acids, \ 6:1,341-344$

To link to this Article: DOI: 10.1080/07328318708056217 URL: http://dx.doi.org/10.1080/07328318708056217

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A NEW 5'-PROTECTING GROUP FOR USE IN SOLID-PHASE SYNTHESIS OF OLIGORIBONUCLEOTIDES

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ABSTRACT. The 2-(2,4-dinitrobenzenesulphenyloxymethyl)benzoyl (DNBSB) group is proposed as a protecting group for the 5'-position of nucleosides. The DNBSB group may be removed under mild non-acidic conditions and may have potential in solid-phase synthesis of oligoribo- and oligodeoxyribonucleotides.

We wish to develop a route for solid-phase synthesis of oligoribonucleotides that involves assembly of nucleotide units in a 3' to 5' direction, similar to that in routine use for oligodeoxyribonucleotides. To this end we have chosen to use acid-labile tetrahydropyranyl (THP) or methoxytetrahydropyranyl (MTHP) for 2'-hydroxyl protection of the nucleotide units and to investigate possible alternative protecting groups for the 5'-position. Such groups must be removed sequentially after each nucleotide addition under conditions where no other protecting group is affected.

We have ruled out the use of acid-labile pixyl or dimethoxytrityl groups for 5'-protection, since we found substantial danger of concomitant loss of THP groups during the sequential acidic treatments required to elaborate an oligonucleotide chain. Instead we have been searching for a 5'-protecting group which can be introduced selectively on to the 5'-position of nucleosides and which can be removed under very mild conditions that do not interfere with oligonucleotide assembly. Although several potentially useful groups have been proposed as 5'-protecting groups one as yet have been reported for use in solid-phase oligoribonucleotide synthesis. We now describe a new 5'-protecting group which may have potential usefulness in solid-phase synthesis.

$$HO \longrightarrow B$$
 (i)
 $HO \cap R$
 (i)
 $HO \cap R$
 (i)
 $HO \cap R$

1 a: R=H . B=T b: R=OR'. R'=THP. B=U c: R=OR'. R'=THP. B=C 2 a; R=H . B=T b; R=OR¹, R¹=THP.B=U c:R=OR¹, R¹=THP.B=C^{B2}

Reagents:(i) DNBSB acid pivaloyl chloride, diisopropylethylamine, dioxane then 4-dimethylaminopyridine, pyridine

FIGURE. Preparation of DNBSB nucleosides

The 2-(2,4-dinitrobenzenesulphenyloxymethyl)benzoyl group (DNBSB) is based on the idea of a 'protected' protecting group originally proposed for nucleosides by Reese^{5,6} but avoids the use of metal ions in the deprotection scheme. The group combines the attributes of the known rapid intramolecularly catalysed deacylation afforded by the 2-hydroxymethylbenzoyl moiety with the 2,4-dinitrobenzenesulphenyl group to protect the hydroxyl function. The preparation of 2-(2,4-dinitrobenzenesulphenyloxymethyl)benzoic acid 10 is accomplished in two steps from phthalide by ring opening with tetra-n-butylammonium hydroxide followed by reaction with 2.4-dinitrobenzenesulphenyl chloride in anhydrous 1,2-dichloroethane. Reaction of thymidine, 2'-0-tetrahydropyranyluridine and 2'-O-tetrahydropyranyl-4-N-benzoylcytidine (la-c) with DNBSB acid together with pivaloyl chloride in anhydrous pyridine yielded after silica gel column chromatography and crystallisation moderate yields (25-33%) of the desired 5'-0-DNBSB nucleosides (2a-c) 10

The DNBSB group is completely removed from nucleosides (2a-c) using 50mM p-toluenethiol and 200mM diisopropylethylamine in acetonitrile/water (90:10 v/v) within 2 minutes. Release of the

2,4-dinitrobenzenesulphenyl group in the first step of the reaction gives rise to a yellow colour which can be quantitated by UV spectroscopy (λ max 456nm).

In order to determine whether these conditions of removal would be compatible with solid-phase synthesis the oligonucleotides, $d[T]_{14}A$ was prepared by the phosphoramidite procedure 11 on a manual flow system, but using 5'-0-pixylthymidine-3'-0-(2-cyanoethyl-N,N-diisopropylamino) phosphite as monomer unit. After each acidic deprotection step the glass support was washed (3 minutes) with 50mM p-toluene thiol and 200mM disopropylethylamine in acetonitrile/water (90:10). High coupling yields were obtained and there was no evidence of significant side reactions. Similarly $d[T]_{6}C$ was prepared by the H-phosphonate procedure 12,13 on a manual flow system but using triethylammonium 5'-0-pixylthymidine-3'-0-H-phosphonate as monomer unit and after each coupling step the intermediate phosphonate was oxidised to the phosphodiester using 2% iodine in pyridine/water (98:2) for 2.5 minutes. Once again after each acidic deprotection step the glass support was washed (3 minutes) with p-toluenethiol and diisopropylethylamine in acetonitrile/water (90:10). Yields were identical to those obtained to a control synthesis without the p-toluenethiol treatment.

Thus far it has not been possible to obtain 5'-0-DNBSB nucleoside 3'-0-(2-cyanoethyl,N,N-diisopropylamino)phosphites in sufficiently pure form. However, we have recently prepared 5'-0-DNBSB-2'-0-tetra-hydropyranyl-4-N-benzoylcytidine-3'-0-H-phosphonate in 74% yield from 1c by the phosphonylation procedure previously described. This phosphonate showed no evidence of decomposition after 24h in pyridine solution as judged by tlc analysis. Attempts at oligonucleotide synthesis using such phosphonates are in progress.

ACKNOWLEDGEMENT

S Agrawal is supported by a scholarship from the Government of India.

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